Cartilage Tissue Engineering Lab



# BACKGROUND

- Measurement of strain in 3D is useful to assess the biomechanical function of tissues and materials [1]
- Micro-computed tomography ( $\mu$ CT) is an x-ray based imaging modality for determining 3D structures with micrometer ( $\mu$ m) level resolutions [2]
- Hydrogels can be created with a variety of material properties, but their deformation and strain behavior under load has previously been studied only in 1-D and 2-D [3]

# HYPOTHESIS

µCT of hydrogels created with radiopaque particles can be analyzed under compression to assess 3D deformation and strain with high resolution.

# **OBJECTIVES**

Create hydrogels with radiopaque particles and evaluate:

- particle size suitable for visualization
- 2. axial and radial deformation and strain
  - A. unconfined compression
  - B. confined compression

# **METHODS**

### **Samples Preparation**

- Gel 3% agarose in Phosphate Buffered Saline (PBS) with 0.1% radiopaque particles
- Disks created: h=3mm, d=6.2mm (Table 1)
- Samples imaged with Skyscan1076 µCT at (9µm)<sup>3</sup> voxel size

### **Samples Compression**

- Samples compressed to 30% over ~150s and 300s, respectively (Fig. 2AB)
- Samples equilibrated for 30 min than imaged in compressed state (Fig. 2CD)

### Image Analysis

- 3D micro CT images thresholded to segment particles from background for multiple grayscale bin ranges
- 3D centroid position of individual particles determined through depth of gel, with z = 0reference located at surface (Fig. 1 A1-3)
- Particle composition determined, with centroid variability between threshold bins For lowest variability particle size, axial and radial displacement of individual particles
- determined, comparing compressed position to initial position
- Displacement vs. position compared and fit to line, with strain being slope **Statistics**
- Standard deviation of particle centroid position for each group
- Standard error for estimate of slopes from regressions (p < 0.05)
- Student's t-test of strain from regression slope and expected imposed strain (p < 0.05)

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Com	Background	Particle	Experimental
	Particle Size (um)	size (um)	Group #
Un	None	NA	1
	20	80	2
	None	80	3
	20	40	4
	None	40	5
С	20	80	6
	None	80	7
	20	40	8
	None	40	9

 
 Table 1. Table of experimental groups tested with particle sizes, different background
attenuation particles and compression type.

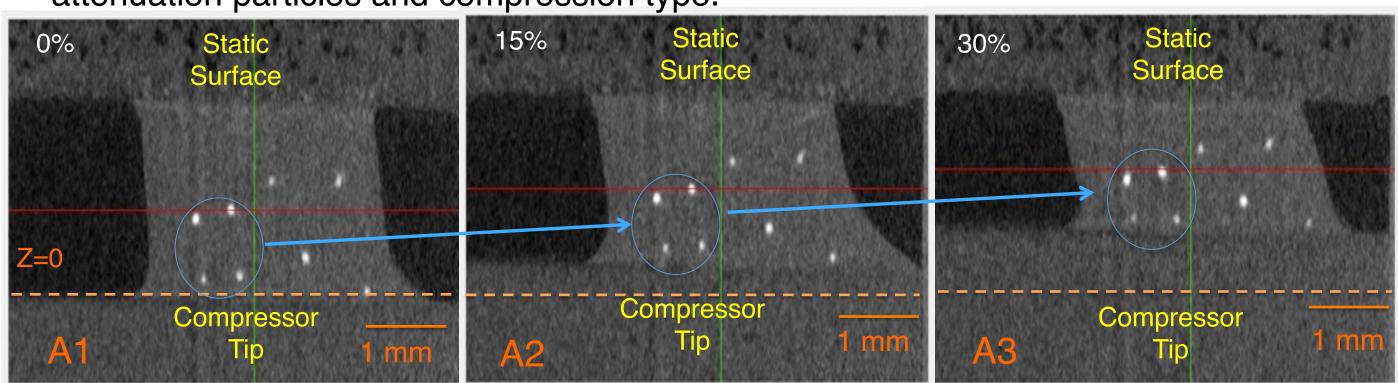


Figure 1. (A1-3) Imaging method of unique groups of particle tracking through 0, 15 and 30% compression (blue arrows).

# Feasibility of 3D Deformation and Strain Analyses by Micro-Computed Tomography Adam Li, Esther Cory, Jason P Caffrey, Van W Wong, Quynhhoa T Nguyen, PhD, Robert L Sah, MD, ScD

Department of Bioengineering & Whitaker Institute of Biomedical Engineering University of California-San Diego, La Jolla, CA

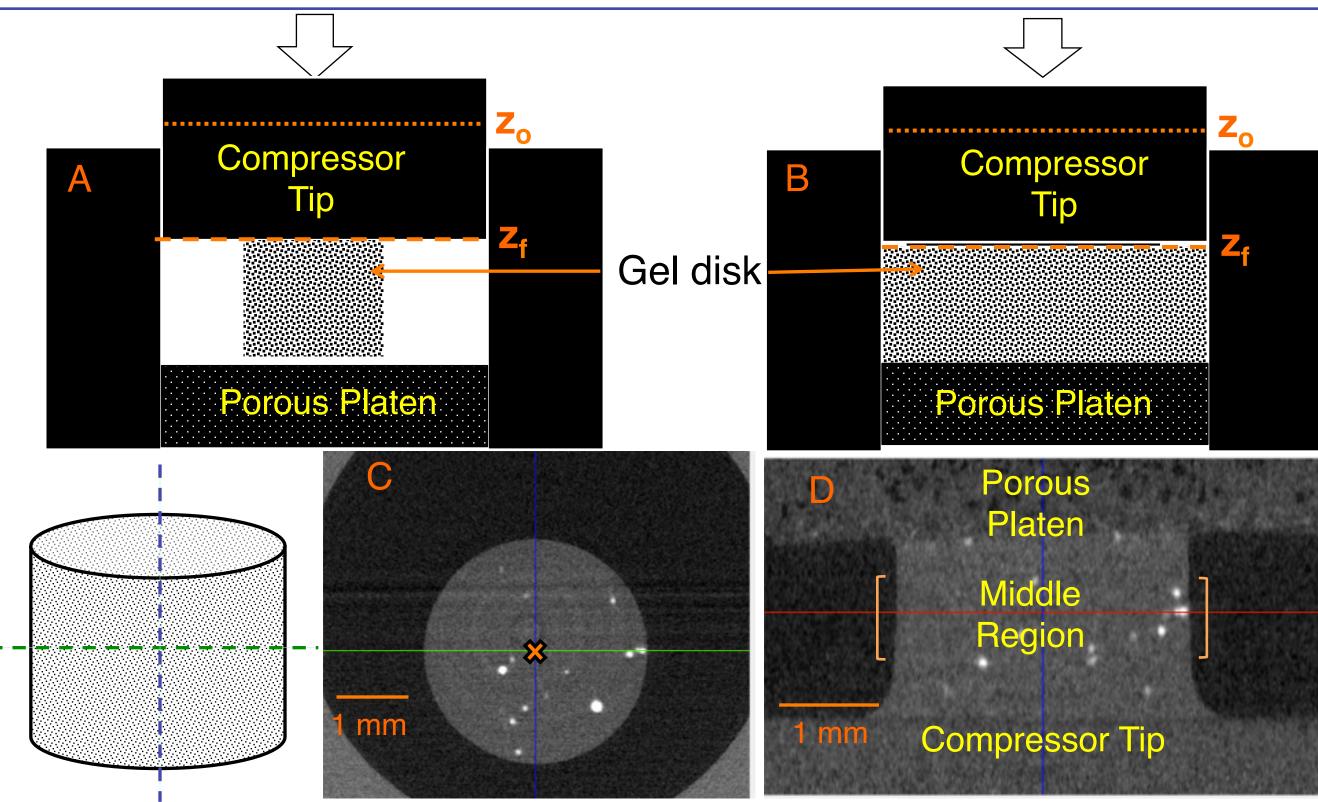


Figure 2. Compression chamber with gel disk in (A) unconfined compression and (B) confined compression. Cross section of hydrogel with the (C) green line showing the orange "X" marks center of the hydrogel used for radial analysis and (D) blue line the middle regions marks area used for radial analysis.

# RESULTS

### **Particle Size and Attenuations**

- (7-9 pixels) (Fig. 3AB)
- 80um particles showed smallest standard deviation of 0.616µm 80µm particle hydrogel without the 20µm background

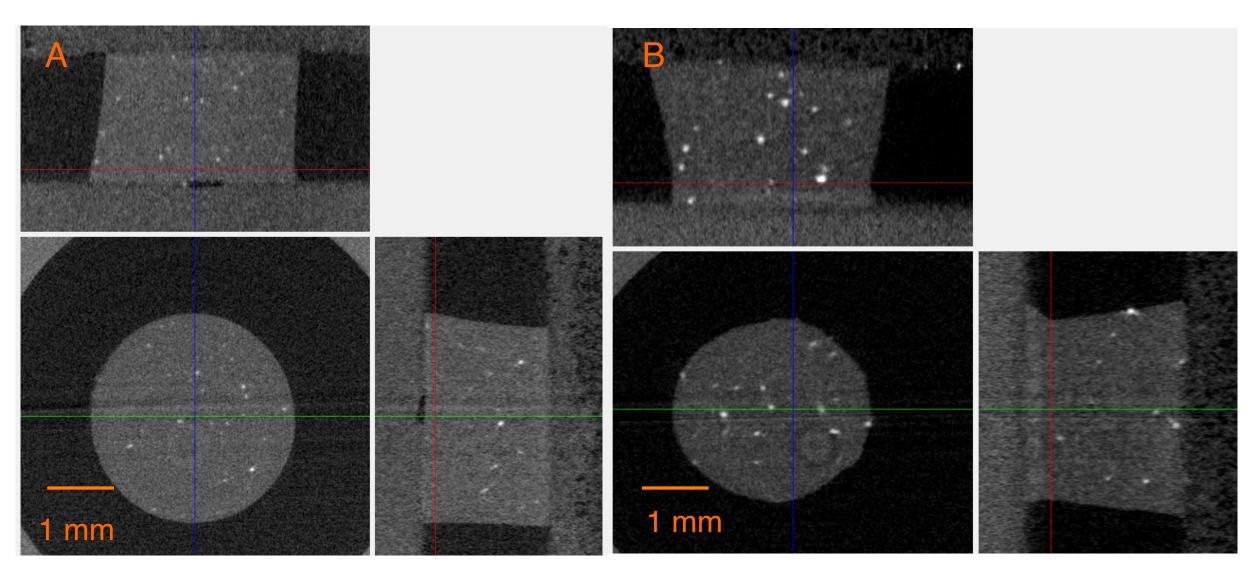
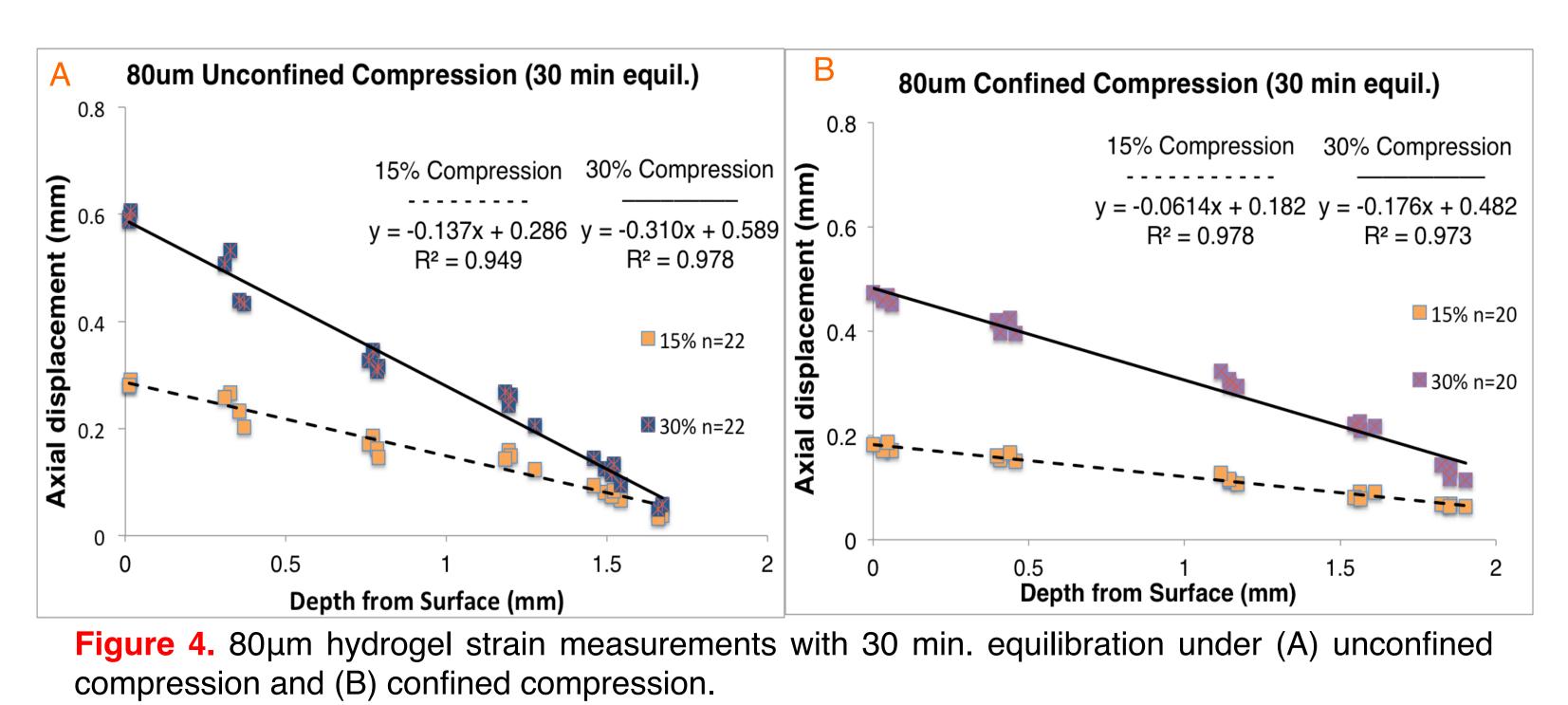


Figure 3. (A) 40µm particle hydrogel shown in orthogonal views and (B) 80µm particle hydrogel shown in orthogonal views.



### mpression type

nconfined

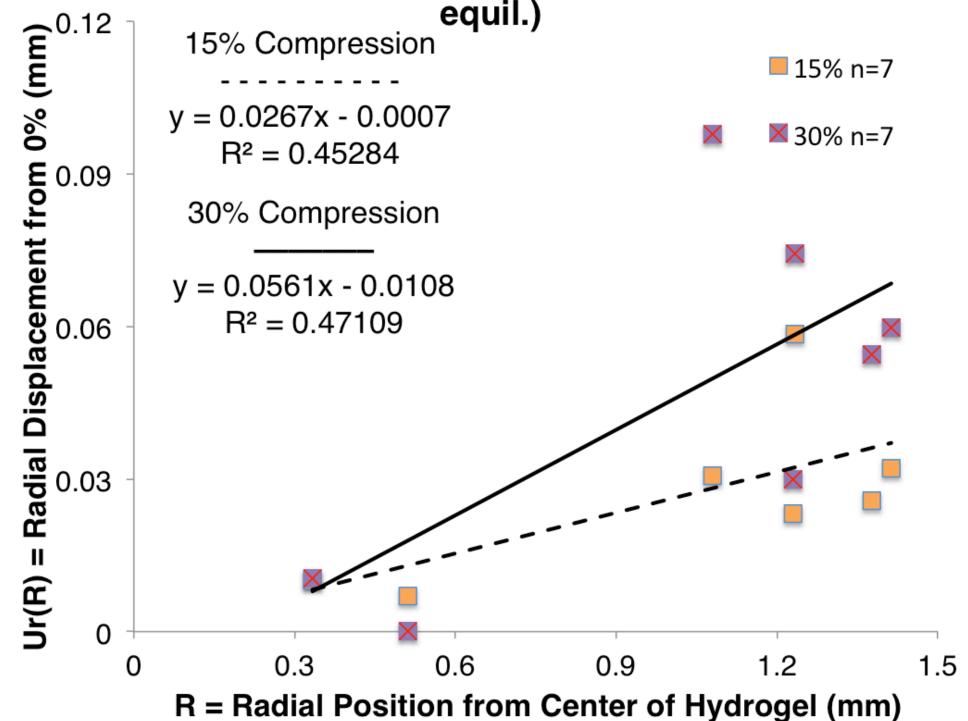
Confined

• Hydrogels containing 40µm particles approached micro CT resolution (2-4 pixels) and were excluded from further analysis, compared to the 80µm particles with much higher resolution

• Adding 20µm particles increased background gel attenuation by 5.3% mm<sup>-1</sup> relative to the

## **Axial and Radial Strain**

- compression, respectively (Fig. 4B)



- feasibility of hydrogel actuator compression
- friction at the confining wall-gel interface
- bulging during unconfined compression

<sup>1</sup>Schinagl RM, Gurskis D, Chen AC, Sah RL. J Orthop Res 1997;15:499-506. <sup>2</sup>Palmer AW, Guldberg RE, Levenston ME.Proc Natl Acad Sci U S A 2006;103:19255-60. <sup>3</sup>Spiller KL, Maher SA, Lowman AM. Tissue Engineering;17:281-299. <sup>4</sup>Mow VC, Kuei SC, Lai WM, Armstrong CG. J Biomech Eng 1980;102:73-84.

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# **RESULTS (Cont.)**

• Linear regression of axial displacement vs. depth for 80um particles yielded a strain of 13.7% and 31.0% for 15% and 30% imposed unconfined compression, respectively (Fig. 4A)

• Linear regressions were statistically significant ( $p \approx 0, 0$ ), R<sup>2</sup>=0.95, 0.98 • Linear regression of axial displacement vs. depth for 80um particles yielded a strain of 6.1% and 17.6% for 15% and 30% imposed confined

Linear regression was statistically significant ( $p \approx 0, 0$ ), R<sup>2</sup>=0.98, 0.97 • Linear regression of radial displacement vs. radius showed low R<sup>2</sup> values of 0.45 and 0.47 for 15% and 30% compression, respectively, which were statistically insignificant (p=0.23, 0.23) (Fig. 5)

# 80um Unconfined Compression (30 min

**Figure 5.** 80µm graph of radial strain with linear regression.

### DISCUSSION

40µm size particles too small for tracking since they approach the micro CT resolution, yielding only 2-4 pixels per particle.

80µm particles had the smallest standard deviation in particle position with different thresholds, and is the best choice for particle tracking

• Adding 20µm increases hydrogel attenuation and may be useful in future studies to distinguishing the hydrogel from surrounding cartilage

Strains for unconfined compression match expected values and demonstrate

Strains for confined compression are lower than expected, which may be due to

Radial strains showed weak linear correlations, which may be due to the movement-dependent method of measurement of the gel's center for the reference position. In future studies, a new origin based on a static point in the image will be used to calculate position and displacement radially.

Other errors as likely due to the skewed nature of the gel and axially uneven

# REFERENCES

### ACKNOWLEDGMENTS